

# The Science of Elastin

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Elastin is an essential part of various human tissues that depend on elasticity. These connective tissues include the skin, lung and arteries. Elastin provides these elastic tissues with the ability to stretch and recoil and plays a critical role in supporting and maintaining healthy cells (Almine et al., 2010; Daamen et al., 2007; Debelle and Alix, 1999; Keeley et al., 2002; Kielty et al., 2002; Mecham, 1991; Pasquali-Ronchetti and Baccarani-Contri, 1997; Pepe et al., 2007; Rosenbloom et al., 1993; Uitto et al., 1991; Vrhovski and Weiss, 1998). In the skin, most elastin is located in the dermis, which is the springy middle layer (Rosenbloom et al., 1993). This elastic tissue is assembled as a continuous network of fibers that encompasses, with decreasing elastin content, the mature elastic fibers, wispy immature elaunin fibers and oxytalan fibers (Montes, 1996). The dense mass of elastic fibers in the reticular dermis dominates the region and is particularly important to the overall elasticity of the skin. Generally, the most mature, thicker elastin fibers are found deep in the dermis, where they function as an inter-penetrating elastin network (Kielty and Shuttleworth, 1997; Ushiki, 2002).

The protein tropoelastin is the fundamental building component of all elastin. There is only one tropoelastin gene (ELN) in humans, in contrast to many other connective tissue proteins like the collagens that can be members of large, complex gene families (Bashir et al., 1989; Indik et al., 1989; Indik et al., 1987). The expression of this single ELN gene mainly occurs before birth and in the first few years of life when the cells of elastic tissues produce the elastin required for the body to develop. From a young age, ELN expression is turned down substantially as we make less elastin, such that by the time we are middle-aged only a trickle of elastin is produced and we rely mostly on the elastin that was

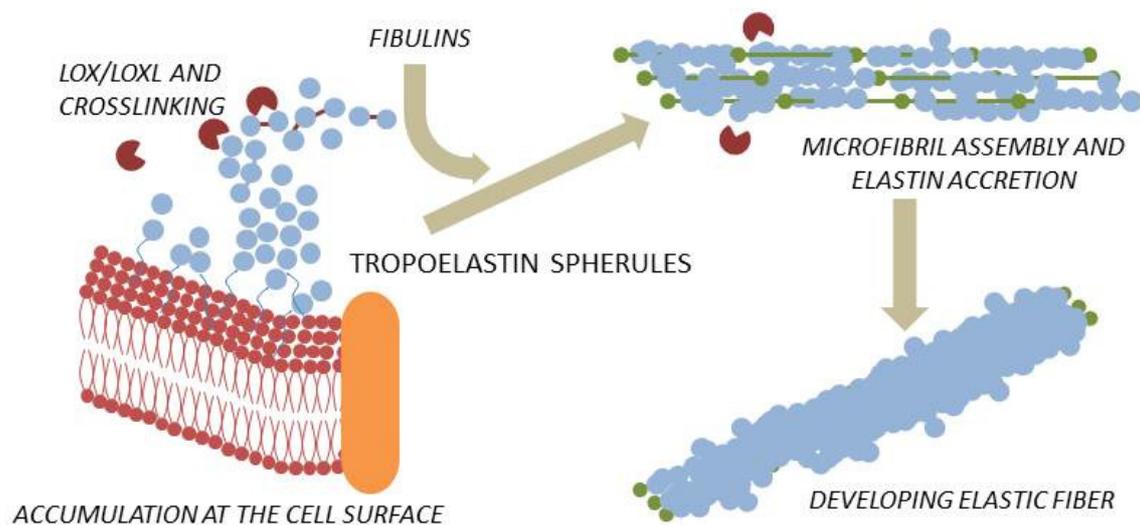
deposited in the womb and those first few years of life (Cleary et al., 1967; Wirtschafter et al., 1967). The implications of this include the fact that our elastic connective tissues need to rely on the persistence of elastin. To this end, Elastin has been shown to have a half-life of about 74 years (Shapiro et al., 1991) and is the longest lasting protein in the body. However, in the event of damage to the elastin in the skin of adults following, e.g., serious injury such as burns; sun damage; or, simply as a result of aging, the low level of elastin production may mean the damage cannot be efficiently repaired and the skin gradually loses its elasticity.

The primary transcript from the single ELN gene is spliced to give different forms of the tropoelastin protein that either lack or contain various exons, which in turn give rise to forms of tropoelastin that can vary slightly in their protein sequence (Indik et al., 1989; Indik et al., 1987). The implications of this splicing variety are not clear although some exons are always present, while others are occasionally spliced out. For example, exon 26A is unique to humans and appears to be spliced out in healthy elastic skin tissue but is occasionally present under conditions of elastin damage, such as following UV exposure (e.g. sun damaged skin) or extreme temperature treatments (Chen et al., 2009; Schmelzer et al., 2005). As such, it may be that some forms of the tropoelastin protein are associated with healthy elastic tissues, while other forms are associated with injury or disease.

As a key step in making elastin, many tropoelastin molecules associate and are then cross-linked, or connected, to form insoluble elastin (Mithieux and Weiss, 2005; Rosenbloom et al., 1993; Vrhovski and Weiss, 1998). The process of elastic fiber formation is also known to include a number of other molecules. The microfibrils, of which fibrillin-1 is the major component, are structures present in the extracellular matrix which are thought to anchor elastic fiber formation. Cross-linking tropoelastin spherules are introduced to the microfibrils by the molecules fibulin-5 and possibly fibulin-4 (Yamauchi et al., 2010) and accrete on pre-existing elastin. Fibulin-2 may work cooperatively with fibulin-5 to assist in elastic fiber formation (Chapman et al., 2010). Emilin-1 may also regulate oxytalan fiber formation but does not appear to directly regulate elastin

expression or deposition (Nakatomi et al., 2011). The cross-linking of tropoelastin is carried out by lysyl oxidases – a family of five enzymes (LOX and LOXL, LOXL 2-4) which are likely to redundantly contribute to the crosslinking process. LOX knockout mice show a reduction in elastin cross-linking (Maki et al., 2005). In addition, LOX and LOXL have both been detected by immunohistochemistry in the dermis and epidermis of normal human foreskin and dermal equivalents and their expression levels have been shown to decrease with age (Noblesse et al., 2004; Sohm et al., 2010).

Figure 1, below, provides a schematic representation of what is currently understood to be the process by which elastin fibers are formed.



**Figure 1: Schematic representation of Elastin fiber formation.** Tropoelastin is expressed then secreted as a mature form of the protein into the extracellular matrix. This tropoelastin accumulates on the cell surface, first as small particles then as larger, approximately 1 micron spherules that are effectively massively associated coacervates of tropoelastin. At an undefined stage, the tropoelastin is subjected to oxidation by lysyl oxidase enzymes at a subset of lysines which subsequently participate in aldol condensation and Schiff base reactions to form cross-links. The forming elastin is introduced to microfibrils in the extracellular matrix by members of the fibulin protein family where the elastin fibers are assembled. The resulting elastin is a very stable and persistent structure that has an impressive ability to confer recoil to human tissues (Debelle and Alix, 1999;

Keeley et al., 2002; Muiznieks et al., 2010; Urry, 1988).

Given the importance of elastin to the skin and its loss in the aging process, it is not surprising that various attempts have been made to maintain or replenish elastin levels. Treatments aiming to repair or regenerate elastin in elastic tissues should consider all the molecules implicated in elastin fiber formation. However, as elastin fibers develop, they ultimately consist of over 90% elastin and so the integration of sufficient tropoelastin into elastin fibers is clearly the major target. Effective treatment approaches are also restricted due to the obvious physical challenge of transferring materials and/or treatments across the epidermis and into the dermis, resulting in a preference for small molecules and physical treatments. Tretinoin or all-trans retinoic acid is a small molecule that has been used for many years in topical formulations to increase elastin production in skin through increased tropoelastin (Bergstrom, 2009; Tajima et al., 1997) and fibrillin expression and secretion (Watson et al., 2008). Molecules such as aldosterone and mineralocorticoid receptor antagonists can impact on elastin fiber deposition in skin (Mitts et al., 2010). Soy (Zhao et al., 2009) and rice (USPTO 19469891) extracts may also increase elastin formation, as can a combination of zinc and copper (Mahoney et al., 2009). Hyperthermia can increase tropoelastin expression and elastic fiber deposition (Murphy et al., 2010) although this may encourage tropoelastin that contains sequences encoded by exon 26A, a region associated with abnormal elastin structures (Chen et al., 2009). When delivered by retroviral overexpression, the extracellular matrix component versican V3 increases tropoelastin expression in disease cells (Hinek et al., 2004). More recently, a dill extract has also been shown to have the potential to promote elastin formation by promoting LOXL synthesis and secretion into the dermis (Cenizo et al., 2006; Sohm et al., 2010).

However, the major challenge which all of the above approaches need to overcome is the very low level of expression of tropoelastin in the adult skin, meaning that such treatments are likely to only have incremental benefits on the density of skin elastin (Sephel et al., 1987).

Researchers have for many years pursued elastin based materials which may be

useful in the repair or regeneration of elastic tissues (Antonicelli et al., 2009; Lupo and Cole, 2007; Zhang and Falla, 2009). Such materials have evolved from fragments of elastin isolated from animal tissue through to synthetic proteins which mimic the elastic nature of tropoelastin. However, to date none of these materials have been capable of truly mimicking the properties of elastin or the tropoelastin building block which combine remarkable physical properties with an ability to support and stimulate the growth of cells present in elastic tissues. Tropoelastin is amongst the most elastic of all known natural proteins (Holst et al., 2010), with an ability to stretch eight times its resting molecular length and recoil without damage to the protein. The adaptation of recombinant gene technology to allow the production of large quantities of clinical grade human tropoelastin, identical to that present in normal human skin, suggests a promising next generation of products for the maintenance and repair of elastin levels in the skin and other elastic tissues is now possible.

## **References**

- Almine, J.F., Bax, D.V., Mithieux, S.M., Nivison-Smith, L., Rnjak, J., Waterhouse, A., Wise, S.G., and Weiss, A.S. (2010). Elastin-based materials. *Chem Soc Rev* 39, 3371-3379.
- Antonicelli, F., Bellon, G., Lorimier, S., and Hornebeck, W. (2009). Role of the elastin receptor complex (S-Gal/Cath-A/Neu-1) in skin repair and regeneration. *Wound Repair Regen* 17, 631-638.
- Bashir, M.M., Indik, Z., Yeh, H., Ornstein-Goldstein, N., Rosenbloom, J.C., Abrams, W., Fazio, M., Uitto, J., and Rosenbloom, J. (1989). Characterization of the complete human elastin gene. Delineation of unusual features in the 5'-flanking region. *J Biol Chem* 264, 8887-8891.
- Bergstrom, K.G. (2009). Beyond tretinoin: cosmeceuticals for aging skin. *J Drugs Dermatol* 8, 674-677.
- Cenizo, V., Andre, V., Reymermier, C., Sommer, P., Damour, O., and Perrier, E. (2006). LOXL as a target to increase the elastin content in adult skin: a dill extract induces the LOXL gene expression. *Exp Dermatol* 15, 574-581.
- Chapman, S.L., Sicot, F.X., Davis, E.C., Huang, J., Sasaki, T., Chu, M.L., and Yanagisawa, H. (2010). Fibulin-2 and fibulin-5 cooperatively function to form the internal elastic lamina and protect from vascular injury. *Arterioscler Thromb Vasc Biol* 30, 68-74.
- Chen, Z., Shin, M.H., Moon, Y.J., Lee, S.R., Kim, Y.K., Seo, J.E., Kim, J.E., Kim, K.H., and Chung, J.H. (2009). Modulation of elastin exon 26A mRNA and protein expression in human skin in vivo. *Exp Dermatol* 18, 378-386.
- Cleary, E.G., Sandberg, L.B., and Jackson, D.S. (1967). The changes in chemical composition during development of the bovine nuchal ligament. *J Cell Biol* 33,

469-479.

Daamen, W.F., Veerkamp, J.H., van Hest, J.C., and van Kuppevelt, T.H. (2007). Elastin as a biomaterial for tissue engineering. *Biomaterials* 28, 4378-4398.

Debelle, L., and Alix, A.J. (1999). The structures of elastins and their function. *Biochimie* 81, 981-994.

Hinek, A., Braun, K.R., Liu, K., Wang, Y., and Wight, T.N. (2004). Retrovirally mediated overexpression of versican v3 reverses impaired elastogenesis and heightened proliferation exhibited by fibroblasts from Costello syndrome and Hurler disease patients. *Am J Pathol* 164, 119-131.

Holst, J., Watson, S., Lord, M.S., Eamegdool, S.S., Bax, D.V., Nivison-Smith, L.B., Kondyurin, A., Ma, L., Oberhauser, A.F., Weiss, A.S., *et al.* (2010). Substrate elasticity provides mechanical signals for the expansion of hemopoietic stem and progenitor cells. *Nat Biotechnol* 28, 1123-1128.

Indik, Z., Yeh, H., Ornstein-Goldstein, N., Kucich, U., Abrams, W., Rosenbloom, J.C., and Rosenbloom, J. (1989). Structure of the elastin gene and alternative splicing of elastin mRNA: implications for human disease. *Am J Med Genet* 34, 81-90.

Indik, Z., Yeh, H., Ornstein-Goldstein, N., Sheppard, P., Anderson, N., Rosenbloom, J.C., Peltonen, L., and Rosenbloom, J. (1987). Alternative splicing of human elastin mRNA indicated by sequence analysis of cloned genomic and complementary DNA. *Proc Natl Acad Sci U S A* 84, 5680-5684.

Keeley, F.W., Bellingham, C.M., and Woodhouse, K.A. (2002). Elastin as a self-organizing biomaterial: use of recombinantly expressed human elastin polypeptides as a model for investigations of structure and self-assembly of elastin. *Philos Trans R Soc Lond B Biol Sci* 357, 185-189.

Kielty, C.M., Sherratt, M.J., and Shuttleworth, C.A. (2002). Elastic fibres. *J Cell Sci* 115, 2817-2828.

Kielty, C.M., and Shuttleworth, C.A. (1997). Microfibrillar elements of the dermal matrix. *Microsc Res Tech* 38, 413-427.

Lupo, M.P., and Cole, A.L. (2007). Cosmeceutical peptides. *Dermatol Ther* 20, 343-349.

Mahoney, M.G., Brennan, D., Starcher, B., Faryniarz, J., Ramirez, J., Parr, L., and Uitto, J. (2009). Extracellular matrix in cutaneous ageing: the effects of 0.1% copper-zinc malonate-containing cream on elastin biosynthesis. *Exp Dermatol* 18, 205-211.

Maki, J.M., Sormunen, R., Lippo, S., Kaarteenaho-Wiik, R., Soinen, R., and Myllyharju, J. (2005). Lysyl oxidase is essential for normal development and function of the respiratory system and for the integrity of elastic and collagen fibers in various tissues. *Am J Pathol* 167, 927-936.

Mecham, R.P. (1991). Elastin synthesis and fiber assembly. *Ann N Y Acad Sci* 624, 137-146.

Mithieux, S.M., and Weiss, A.S. (2005). Elastin. *Adv Protein Chem* 70, 437-461.

Mitts, T.F., Bunda, S., Wang, Y., and Hinek, A. (2010). Aldosterone and mineralocorticoid receptor antagonists modulate elastin and collagen deposition

- in human skin. *J Invest Dermatol* 130, 2396-2406.
- Montes, G.S. (1996). Structural biology of the fibres of the collagenous and elastic systems. *Cell Biol Int* 20, 15-27.
- Muiznieks, L.D., Weiss, A.S., and Keeley, F.W. (2010). Structural disorder and dynamics of elastin. *Biochem Cell Biol* 88, 239-250.
- Murphy, B.A., Bunda, S., Mitts, T., and Hinek, A. (2010). The hyperthermia-enhanced association between tropoelastin and its 67-kDa chaperone results in better deposition of elastic fibers. *J Biol Chem* 285, 40282-40293.
- Nakatomi, Y., Tsuruga, E., Nakashima, K., Sawa, Y., and Ishikawa, H. (2011). EMILIN-1 regulates the amount of oxytalan fiber formation in periodontal ligaments in vitro. *Connect Tissue Res* 52, 30-35.
- Noblesse, E., Cenizo, V., Bouez, C., Borel, A., Gleyzal, C., Peyrol, S., Jacob, M.P., Sommer, P., and Damour, O. (2004). Lysyl oxidase-like and lysyl oxidase are present in the dermis and epidermis of a skin equivalent and in human skin and are associated to elastic fibers. *J Invest Dermatol* 122, 621-630.
- Pasquali-Ronchetti, I., and Baccarani-Contri, M. (1997). Elastic fiber during development and aging. *Microsc Res Tech* 38, 428-435.
- Pepe, A., Bochicchio, B., and Tamburro, A.M. (2007). Supramolecular organization of elastin and elastin-related nanostructured biopolymers. *Nanomed* 2, 203-218.
- Rosenbloom, J., Abrams, W.R., and Mecham, R. (1993). Extracellular matrix 4: the elastic fiber. *Faseb J* 7, 1208-1218.
- Schmelzer, C.E., Getie, M., and Neubert, R.H. (2005). Mass spectrometric characterization of human skin elastin peptides produced by proteolytic digestion with pepsin and thermolysin. *J Chromatogr A* 1083, 120-126.
- Sephel, G.C., Buckley, A., and Davidson, J.M. (1987). Developmental initiation of elastin gene expression by human fetal skin fibroblasts. *J Invest Dermatol* 88, 732-735.
- Shapiro, S.D., Endicott, S.K., Province, M.A., Pierce, J.A., and Campbell, E.J. (1991). Marked longevity of human lung parenchymal elastic fibers deduced from prevalence of D-aspartate and nuclear weapons-related radiocarbon. *J Clin Invest* 87, 1828-1834.
- Sohm, B., Cenizo, V., Andre, V., Zahouani, H., Pailler-Mattei, C., and Vogelgesang, B. (2010). Evaluation of the efficacy of a dill extract in vitro and in vivo. *Int J Cosmet Sci*.
- Tajima, S., Hayashi, A., and Suzuki, T. (1997). Elastin expression is up-regulated by retinoic acid but not by retinol in chick embryonic skin fibroblasts. *J Dermatol Sci* 15, 166-172.
- Uitto, J., Christiano, A.M., Kahari, V.M., Bashir, M.M., and Rosenbloom, J. (1991). Molecular biology and pathology of human elastin. *Biochem Soc Trans* 19, 824-829.
- Urry, D.W. (1988). Entropic elastic processes in protein mechanisms. II. Simple (passive) and coupled (active) development of elastic forces. *J Protein Chem* 7, 81-114.

Ushiki, T. (2002). Collagen fibers, reticular fibers and elastic fibers. A comprehensive understanding from a morphological viewpoint. *Arch Histol Cytol* 65, 109-126.

Vrhovski, B., and Weiss, A.S. (1998). Biochemistry of tropoelastin. *Eur J Biochem* 258, 1-18.

Watson, R.E., Long, S.P., Bowden, J.J., Bastrilles, J.Y., Barton, S.P., and Griffiths, C.E. (2008). Repair of photoaged dermal matrix by topical application of a cosmetic 'antiageing' product. *Br J Dermatol* 158, 472-477.

Wirtschafter, Z.T., Cleary, E.G., Jackson, D.S., and Sandberg, L.B. (1967). Histological changes during the development of the bovine nuchal ligament. *J Cell Biol* 33, 481-488.

Yamauchi, Y., Tsuruga, E., Nakashima, K., Sawa, Y., and Ishikawa, H. (2010). Fibulin-4 and -5, but not Fibulin-2, are Associated with Tropoelastin Deposition in Elastin-Producing Cell Culture. *Acta Histochem Cytochem* 43, 131-138.

Zhang, L., and Falla, T.J. (2009). Cosmeceuticals and peptides. *Clin Dermatol* 27, 485-494.

Zhao, R., Bruning, E., Rossetti, D., Starcher, B., Seiberg, M., and Iotsova-Stone, V. (2009). Extracts from Glycine max (soybean) induce elastin synthesis and inhibit elastase activity. *Exp Dermatol* 18, 883-886.